

Mitochondrial genome sequencing in phenotype-based panels and exome sequencing increases test sensitivity

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Introduction

Next-generation sequencing (NGS) allows rapid variant analysis for identifying disease causing variants. Exclusion of mitochondrial sequencing analysis in NGS disease screening panels limits their power by ignoring the potential of pathogenic mitochondrial variants. Initially thought to be rare, mitochondrial genetic diseases represent important and common sources of disorders. Recent genetic epidemiological studies quantifying the most common pathogenic mtDNA variants have shown the incidence of clinical mitochondrial diseases is about 1 in 5000¹. Additionally, a survey of newborn cord bloods revealed that 1 in 200 infants harbored common pathogenic mtDNA variants^{2,3}. Coupled with variant assessment and curation database software, the addition of mtDNA sequencing to phenotypebased panels and exomes increases the ability to discover variants of interest that aid in the determination of patient treatment. Upon including mtDNA sequencing with the panels, we have positively identified pathogenic or potentially pathogenic variants in patient samples having no definitively pathogenic nuclear genome variants. The effect of this inclusion has been particularly beneficial in NGS panels associated with neurological disorders. The increased sensitivity of the NGS panels and the ease of ordering a single test instead of two provides added value for both clinicians and their patients.

Methodology

 MITOCHONDRIAL DNA SINGLE NUCLEOTIDE POLYMORPHISM, SMALL INDEL SEQUENCING, AND DELETION ASSESSMENT

Next-generation sequencing of a long-range mtDNA PCR product of approximately 16,476 base pairs is performed on the Illumina® MiSeq™ instrument. This region encompasses all known pathogenic variants in the mtDNA. Deletion assessment is performed based on identification of regions flanking a deletion (LUMPY⁴).

• RESULTS INTERPRETATION

Interpretation of PATHOGENIC and LIKELY PATHOGENIC variants, as well as Variants of Uncertain Significance (VUS) with pathogenic predictions or possible disease association related to the patient's phenotype are determined using online database resources (e.g., ClinVar⁵, OMIM⁶, MitoMap⁷) and our inhouse Genome MaNaGer® variant curation database software.

PANELS WITH mtDNA ANALYSIS INCLUDED

The data reported were collected from the following NGS panels which include mtDNA sequencing and deletion

analysis:

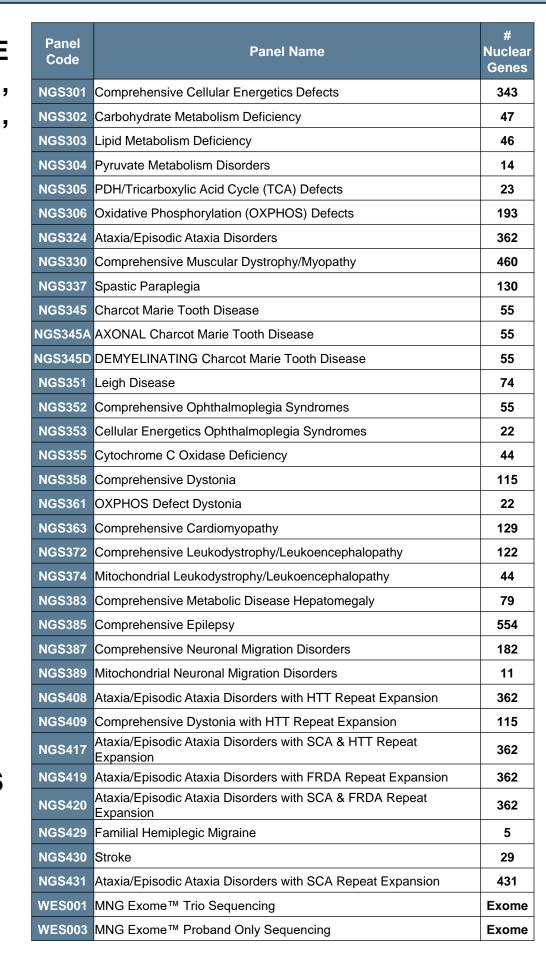
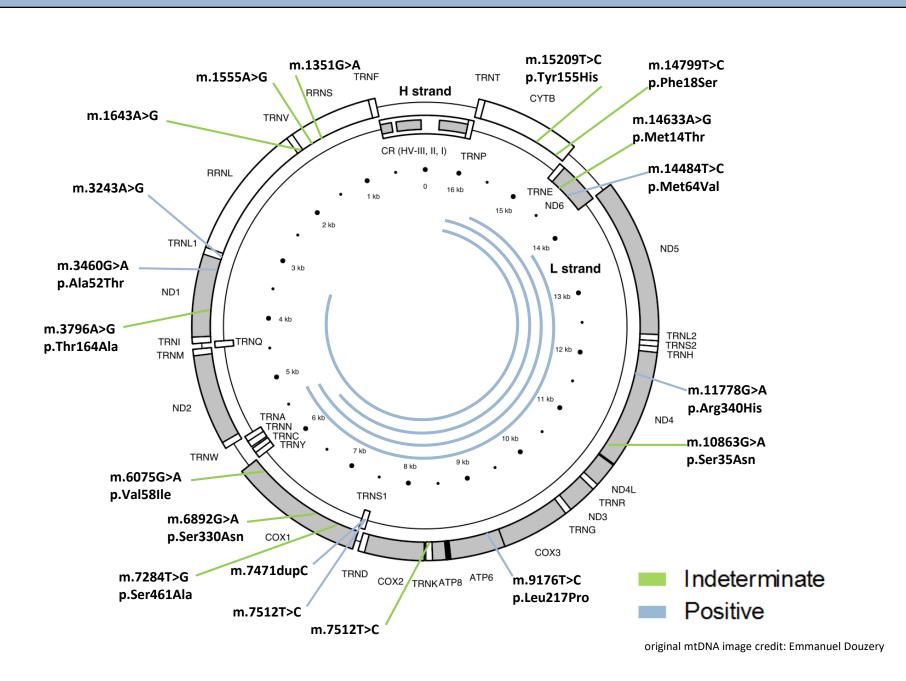


FIG 2: Reported mtDNA variants

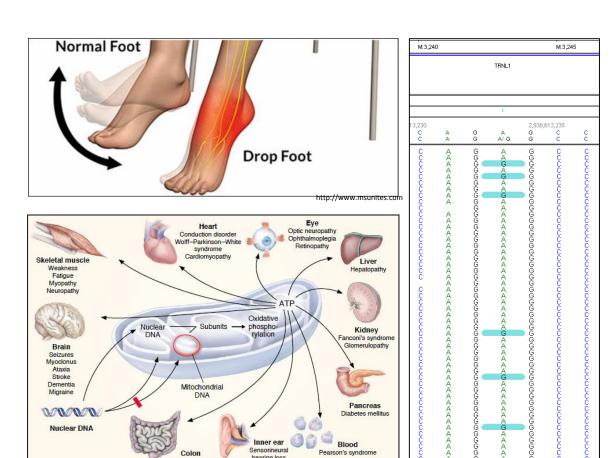


Variants shown were responsible for either Indeterminate reports or Positive reports of NGS panels with mtDNA sequencing included. Inner curved lines represent approximate lengths and locations of reported deletions.

FIG 3: MELAS detection in exome

Clinical Information

- 14 year-old male
- Progressive gait abnormality of unknown etiology
- Proximal limb weakness and areflexia
- Bilateral foot drops



Leukocyte sample from exome patient determined to be at 11% heteroplasmy for the m.3243A>G MELAS variant (raw data on left). Clinician stated discovery of MELAS was unexpected but was a fitting diagnosis for the clinical presentation of the patient (clinical information above).

Standard exomes excluding mtDNA sequencing would not have provided diagnostic evidence for cause of disease in this case.

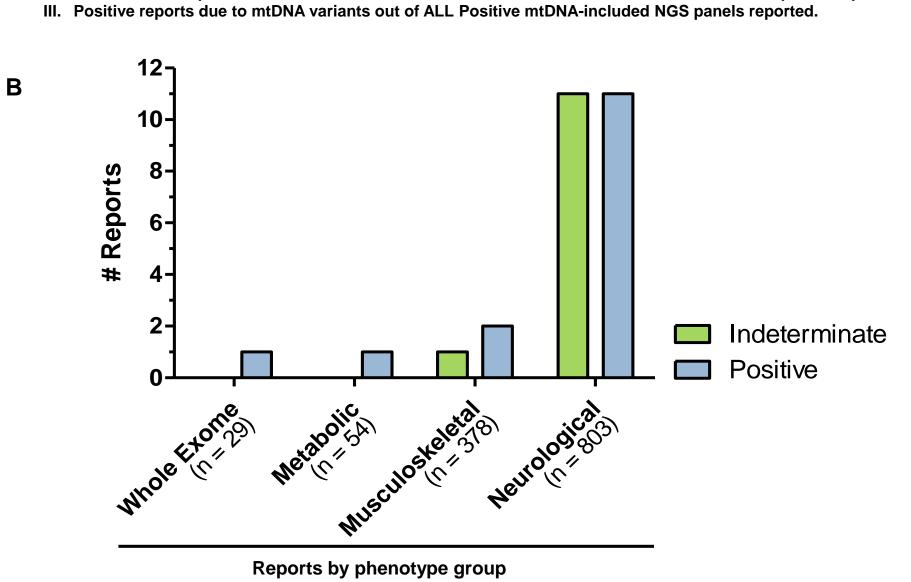
TABLE 1: Reported Variant Details

Reported Variant (Positive Reports)	Panel Description	Variant Info with Heteroplasmy Percentage (DNA from leukocytes unless specified)	Patient Phenotype from Clinical	Age
mtDNA deletion (m.3273-16072)	NGS352 Comprehensive Ophthalmoplegia Syndromes	mtDNA deletion, in 34% of the mitochondria. (frozen muscle)	Ptosis, imbalance, muscle weakness, numbness to lower legs and hands, diabetes mellitus, obesity.	62
mtDNA deletion (m.5841-16070)	NGS352 Comprehensive Ophthalmoplegia Syndromes	mtDNA deletion, in 9% of the mitochondria. (frozen muscle)	No clinical information was provided.	64
ntDNA deletion (m.6340-13994)	NGS352 Comprehensive Ophthalmoplegia Syndromes	mtDNA deletion, in 10% of the mitochondria. (frozen muscle)	CPEO, ptosis, fatigue, limb weakness, migraines, cardiomegaly, right sided heart failure, lipomas, short stature, depression, cerebral atrophy, low CPK.	56
ntDNA deletion (m.6469-15591)	NGS301 Comprehensive Cellular Energetics Defects	mtDNA deletion, in 65% of the mitochondria.	Mitochondrial myopathy.	10
1T-TL1 m.3243A>G	NGS330 Comprehensive Muscular Dystrophy/Myopathy	MELAS @ 2.7%	Proximal lower extremity weakness, normal CPK.	12
/IT-TL1 m.3243A>G	NGS330 Comprehensive Muscular Dystrophy/Myopathy	MELAS @ 1.4%	Myopathy.	88
MT-TL1 m.3243A>G	NGS385 Comprehensive Epilepsy	MELAS @ 17.4%	No clinical information was provided.	1
IT-TL1 m.3243A>G	NGS372 Comprehensive Leukodystrophy/Leukoencephalopathy	MELAS @ 32%	No clinical information was provided.	53
IT-TL1 m.3243A>G	WES001 MNG Exome™ Trio	MELAS @ 11%	Progressive gait abnormality of unknown etiology, limb weakness, areflexia, bilateral foot drops.	14
T-ND1 m.3460G>A Ala52Thr	NGS324 Ataxia/Episodic Ataxia Disorders	LHON @ 16%	Hereditary ataxia	3
IT-TS1 m.7471dupC	NGS385 Comprehensive Epilepsy	PEM/AMDF/Motor neuron disease-like @ 25%	Focal seizures, EMG/NCV show generalized spikes, 1-6x focal seizures per day, eye deviation to left side, negative family history.	2
MT-TS1 m.7512T>C	NGS385 Comprehensive Epilepsy	MELAS/MERFF overlap @ 8.1%	Tonic clonic seizures, stroke, EEG shows focal slowing, family history of epilepsy.	13
MT-ATP6 m.9176T>C Leu217Pro	NGS324 Ataxia/Episodic Ataxia Disorders	Leigh / Bilateral striatal necrosis @ 96% (extracted DNA, tissue not specified)	Autosomal recessive spastic ataxia.	51
//T-ND4 m.11778G>A Arg340His	NGS372 Comprehensive Leukodystrophy/Leukoencephalopathy	LHON - homoplasmic	Leukoencephalopathy and neonatal onset seizures of uncertain etiology, controlled with medication.	1
IT-ND6 m.14484T>C Met64Val	NGS324 Ataxia/Episodic Ataxia Disorders	LHON - homoplasmic	No clinical information was provided.	2
11 NDO III.14404170 IIICCO4Vai	1100024 / Maxia/Episodio / Maxia Disorders	El 1014 Homopiasinic	ivo cilinical il normation was provided.	
Reported Variants (Indeterminate Reports)	Panel Description	Variant Info with Heteroplasmy Percentage (DNA from leukocytes unless specified)	Patient Phenotype from Clinical	Age
IT-RNR1 1351G>A	NGS324 Ataxia/Episodic Ataxia Disorders	VUS @ 83.5% . Variants in this gene have been seen in cases of deafness, cardiomyopathy.	No clinical information was provided.	84
MT-RNR1 1555A>G	NGS372 Comprehensive Leukodystrophy/Leukoencephalopathy	Homoplasmic. Variant is known to cause deafness due to use of aminoglycoside antibiotics.	Degenerative disease of nervous system, unspecified, no deafness reported.	5
T-TV m.1643A>G	NGS385 Comprehensive Epilepsy	VUS, homoplasmic. Variant seen in patient with symptoms of epileptic status with myoclonic jerks, but mother was 60% heteroplasmic.	No clinical information was provided.	9
/IT-ND1 m.3796A>G Thr164Ala	NGS385 Comprehensive Epilepsy	VUS, homoplasmic. Reported in publication of patient with adult-onset dystonia.	Localization-related idiopathic epilepsy and epileptic syndromes with seizures of localized onset, not intractable, without status epilepticus, EEG shows left centrotemporal sharp waves, tonic clonic epilepsy, maternal great uncle with Lennox-Gastaut Syndrome, mother has seizures, father hydrocephalus, healthy brother.	8
MT-CO1 m.6075G>A Val58lle	NGS330 Comprehensive Muscular Dystrophy/Myopathy	VUS, homoplasmic in leukocytes. Variants in this gene have been seen in cases of cytochrome C oxidase deficiency, sideroplastic anemia, optic atrophy, deafness, and colorectal cancer.	No clinical information was provided.	28
MT-CO1 m.6892G>A Ser330Asn	NGS358 Comprehensive Dystonia	VUS, homoplasmic in DNA sample. Variants in this gene have been seen in cases of cytochrome C oxidase deficiency, sideroplastic anemia, optic atrophy, deafness, and colorectal cancer. (extracted DNA, tissue not specified)	Intermittent jerky movements of the right upper limb and neck, movements are episodic, quick and jerky, brain MRI shows diffuse increase in the thickness of the posterior body and the splenium of the corpus callosum, no jaw tensing, no head or hand tremors, parents are distantly related, 6 healthy siblings.	13
MT-CO1 m.7284T>G Ser461Ala	NGS345A AXONAL Charcot Marie Tooth Disease	VUS, homoplasmic in leukocytes. Variants in this gene have been seen in cases of cytochrome C oxidase deficiency, sideroplastic anemia, optic atrophy, deafness, and colorectal cancer.	Idiopathic progressive neuropathy.	52
MT-TK m.8296A>G	NGS372 Comprehensive Leukodystrophy/Leukoencephalopathy	VUS, homoplasmic. Variant seen in patients with MELAS, optic atrophy, bilateral striatal necrosis, epilepsy, cardiomyopathy, stroke-like episodes, and diabetes. Cybrid studies show variant may cause defects in oxidative phosphorylation.	Spastic diplegic cerebral palsy, congenital hypotonia, intellectual disability, staring spells, normal EEG, MRI during infancy showed abnormal white matter changes not present in recent MRI, hypotonia in upper extremities, obstructive sleep apnea, normal microarray.	12
MT-ND4 m.10863G>A Ser35Asn	NGS372 Comprehensive Leukodystrophy/Leukoencephalopathy	VUS, homoplasmic. Variants in this gene have been seen in cases of optic atrophy, MELAS, dystonia, complex 1 deficiency.	Atypical febrile seizures, mild speech delay, abnormal brain MRI, T2 flair signal in the centrum semiovale and periventricular white matter and the splenium of the corpus collosum with enlarged perivascular spaces predominantly in white matter.	5
	NGS385 Comprehensive Epilepsy	VUS, homoplasmic. Variants in this gene have been seen in cases of optic atrophy, MELAS, dystonia, complex 1 deficiency, bilateral striatal necrosis, oncocytoma, Parkinson's.	No clinical information was provided.	3
MT-ND6 m.14633A>G Met14Thr				1
MT-ND6 m.14633A>G Met14Thr MT-CYB m.14799T>C Phe18Ser	NGS372 Comprehensive Leukodystrophy/Leukoencephalopathy	VUS @ 13%. Variants in this gene have been seen in cases of optic atrophy, exercise intolerance, encephalomyopathy, multisystem disorder, septooptic dysplasia, obesity, and parkinsonism/MELAS overlap syndrome. VUS, homoplasmic. Previously seen in a patient with Prader-Willi with multisystem	Tonic clonic seizures, developmental delay, ocular apraxia, parents and sibling are healthy, negative family history.	5

FIG 1: Number of reported mtDNA variants

Α		Data Range: Oct 2016-Aug 2017	Totals
	I.	# NGS Panel Tests with mtDNA	1265
		% All NGS Panel Indeterminate Tests due to mtDNA	12/1265 = 0.9%
		% All NGS Panel Positive Tests due to mtDNA	15/1265 = 1.2%
	II.	# Indeterminate NGS Panel Tests with mtDNA	733
		% NGS Panel Indeterminate Tests due to mtDNA	12/733 = 1.6%
	III.	# Positive NGS Panel Tests with mtDNA	232
		% NGS Panel Positive Tests due to mtDNA	15/232 = 6.5%

Indeterminate and Positive reports due to mtDNA variants out of ALL mtDNA-included NGS panels reported.
 Indeterminate reports due to mtDNA variants out of ALL Indeterminate mtDNA-included NGS panels reported.



Results

- mtDNA variants were responsible for 6.5% of all positive reports in panels screening for both nuclear and mitochondrial genes (Figure 1A).
 Additionally, neurological phenotype-based panels were the group with the highest number of mtDNA variants of interest (Figure 1B).
- Reported mtDNA variants were detected throughout the mitochondrial chromosome in protein-coding, tRNA, and rRNA genes. Large mitochondrial deletions were also detected. (Figure 2 and Table 1).
- Inclusion of mtDNA in exome sequencing allowed for identification of the m.3243A>G MELAS variant in a patient whose symptoms were not initially suspected to be of mitochondrial origin (Figure 3).

Conclusions

- Mitochondrial dysfunction is important in pathogenesis and clinical manifestation of disease^{8,9} making mtDNA analysis both a logical and practical addition for sequencing panels.
- Benefits to including mtDNA analysis not only manifest in the identification of variants of interest that could direct patient care, but also in relation to the cost, time, and interpretation that would be required for ordering and receiving separate results for individual nuclear and mitochondrial sequencing panels. This is especially important in cases where the rapid decline of a patient is of concern.
- Our data demonstrate these benefits, through identification of pathogenic variants in patients that would have gone undetected in nuclear gene only testing. Increased sensitivity to identify variants of interest, combined with the efficiency of ordering a single test, provides added value and increases the potential of proper clinical diagnoses.

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